## Stereochemistry of the Decarboxylation of Phenolic Cinnamic Acids by Saccharomyces cerevisiae

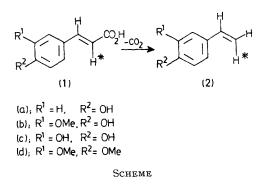
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Summary The decarboxylation of (E)-3,4-dimethoxycinnamic acid to the corresponding styrene by S. cerevisiae proceeds with retention of the hydrogen atom at the  $\alpha$ -position of the acid; the configuration of the double bond is also retained.

ALTHOUGH the non-oxidative decarboxylation of phenolic cinnamic acids [e.g. p-coumaric (1a), ferulic (1b), and caffeic (1c) acids] to the corresponding styrenes (2) occurs widely in bacteria<sup>1,2</sup> and in yeast,<sup>1,3</sup> the enzymic mechanism is unknown. As a preliminary approach to its clarification, the stereochemical course of the reaction has been investigated. We now report that the decarboxylation of 3,4-dimethoxycinnamic acid (1d) by a strain of Saccharomyces cerevisiae<sup>†</sup> results in retention of configuration at the side-chain double bond (Scheme).

A culture medium<sup>‡</sup> containing a suspension of (E)-3,4dimethoxy[a-<sup>2</sup>H]cinnamic acid (1d, H<sup>\*</sup> = <sup>2</sup>H) (D atoms per molecule  $0.80 \pm 0.03$  by m.s.)<sup>4</sup> was inoculated with yeast and shaken at 25° for 24 h. The ether extract of the fermentation medium, when evaporated and chromatographed on silica gel [light petroleum (b.p. 40-70°)benzene 1:1], gave (Z)-3,4-dimethoxy[ $\beta$ -<sup>2</sup>H]styrene (2d, H\* = <sup>2</sup>H)<sup>5</sup> (62% yield; D atoms per molecule 0.75 ± 0.03



† This strain (28 C)<sup>3</sup> is unique in decarboxylating both 3,4-dimethoxycinnamic acid and ferulic acid. Its use made our investigation easier, 3,4-dimethoxystyrene being more stable than 4-hydroxystyrenes.

 $\ddagger$  Glucose (100 g), yeast nitrogen base (Difco) (7 g), NaH<sub>2</sub>PO<sub>4</sub> (13 g), water (1 l).

by m.s.). The position of the deuterium atom was assigned by comparison of the <sup>1</sup>H n.m.r. spectrum (vinyl group region) of (2d) with the spectral patterns calculated for each of the three 3,4-dimethoxystyrenes monodeuteriated in their side-chain. The calculations, (using the secondorder perturbation method), were based on the chemical shifts and spin-coupling constants of the vinyl protons of 3,4-dimethoxystyrene ( $\delta_A$  5.56,  $\delta_B$  5.12,  $\delta_X$  6.66 p.p.m.;  $J_{AX}$  17.5,  $J_{BX}$  10.6,  $J_{AB}$  1.4 Hz in CDCl<sub>3</sub>) and assumed (a) that  $J_{\rm HD} = (\gamma_{\rm D}/\gamma_{\rm H}) J_{\rm HH}$  and (b) that the chemical shifts are not affected by deuterium substitution.6 The correctness of these assumptions was confirmed by comparing the observed and theoretical spectrum of (E)-3,4-dimethoxy- $[\beta^{-2}H]$  styrene prepared by unequivocal synthesis via  $D_2O$ decomposition of the Grignard reagent<sup>6</sup> of trans-3,4-dimethoxy- $\beta$ -bromostyrene.<sup>7</sup>

If the hypothesis is made that the *in vivo* decarboxylation of cinnamic acids takes place similarly to the in vitro pyridine or thioacetic acid-catalysed decarboxylation of benzylidenemalonic acid derivatives,<sup>8</sup> i.e. by a 1,2-addition,

1,2-elimination mechanism (equation 1), then a *cis*-addition followed by a trans-decarboxylative elimination (or a trans-addition and cis-elimination) must be assumed to account for the overall stereochemistry of the process.

$$Ar-CH_{2}CH_{2}-CO_{2}H \xrightarrow{+x^{-},+H^{+}} Ar-CH_{2}CH_{2}-CO_{2}H \xrightarrow{-x^{-},-H^{+},-CO_{2}} Ar-CH_{2}(1)$$

 $(X^- = nucleophilic group of the enzyme, e.g. RS^-, RO^-)$ 

It is also remarkable that (Z)-3,4-dimethoxycinnamic acid<sup>9</sup> does not undergo decarboxylation by the above strain of S. cerevisiae.

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 R. D. Steinke and M. C. Paulson, J. Agric. Food Chem., 1964, 12, 381.
 S. R. Indahl and R. R. Scheline, Appl. Microbiol., 1968, 16, 667; B. J. Finkle, J. C. Lewis, J. W. Corse, and R. E. Lundin, J. Biol. Chem., 1962, 237, 2926

<sup>3</sup> G. Albagnac and P. Dubois, Les phénols volatils des milieux fermentés in 'Comptes rendus de l'Assemblée 1972 du Groupe Poly-phénols,' Station de Technologie des Produits Vegetaux (I.N.R.A.), Narbonne, 1973.

P. Manitto, D. Monti, P. Gramatica, and E. Sabbioni, J.C.S. Chem. Comm., 1973, 563.
 G. Redeuilh, P. Rumpf, and C. Viel, Bull. Soc. chim. France, 1973, 2665.
 T. Yoshino, Y. Manabe, and Y. Kikuchi, J. Amer. Chem. Soc., 1964, 86, 4670.

<sup>7</sup> E. Adler and K. J. Björkqvist, Acta Chem. Scand., 1951, 5, 241; E. R. Trumbull, R. T. Finn, K. M. Ibne-Rasa, and C. K. Sauers, J. Org. Chem., 1962, 27, 2339.
<sup>8</sup> E. J. Corey and G. Fraenkel, J. Amer. Chem. Soc., 1953, 75, 1168.
<sup>9</sup> E. Adler and B. Gustafsson, Acta Chem. Scand., 1963, 17, 27.